

Serological analysis for torch infection in women with bad obstetric record

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ABSTRACT

Infant death within utero or at delivery has always been a disgraceful occurrence for perinatal death and remains a dispute in the concern of pregnant women worldwide. Bad Obstetric record implies consecutive spontaneous abortions, history of intrauterine fetal death, intrauterine growth retardation or congenital abnormalities. The purpose of the study is to reveal the evidence of TORCH infections among pregnancy wastage in women which had bad obstetric history and the study also assessed the sero prevalence of TORCH infection in antenatal women of correlated with BOH by using NANO ELISA method. Blood samples for this study were collected from different hospitals in and around Bangalore and Bagalkot city of Karnataka state. From the study it was evident that those maternal infections play a critical role in pregnancy wastage and their occurrence in the patients with BOH is a significant factor. Usage of NANO ELISA technique provides high sensitive, accurate results at economical price.

Key words: Torch, Nano Elisa, Bad Obstetric History

1. INTRODUCTION

Bad obstetric history (BOH) implies previous unfavorable fetal outcome in terms of two or more consecutive spontaneous abortions, history of intrauterine foetal death, intrauterine growth retardation, stillbirth, early neonatal death and/or congenital anomalies. Infections by TORCH agents in women are usually asymptomatic and chronic. Fetus TORCH infections cause a syndrome characterized by microcephaly, sensorineural deafness, chorioretinitis, hepatosplenomegaly and thrombo-cytopenia. Agglutination tests are easy to

perform for serological observation. Rubella is a well-known as German Measles a viral disease which characterized by patent ductus arteriosus, septal defects, pulmonary artery stenosis, sensory-neural deafness, meningoencephalitis, Intrauterine growth retardation (IUGR), and osseous changes. Less than 5% of pregnant women with primary infection are reported to be symptomatic, and even smaller percentages suffer from a mononucleosis syndrome. Herpes simplex virus (HSV) commonly causes infections of the skin and mucous membranes.

The Present study was aimed to reveal the incidence of ToRCH infections among pregnancy wastage in women which had Bad Obstetric History. This study is also conducted to assess the Seroprevalence of ToRCH infection in antenatal women and correlate with Bad Obstetric History by using nanoelisa method. ToRCH describes a group of clinically similar congenital infections caused by Toxoplasma gondii, Rubella, CMV and HSV1 and 2. Nanowells are much smaller in size (Cubic volume) in comparison to conventional ELISA microwells. As a result, NanoWells require less reagents and are highly cost efficient. Unlike conventional ELISA, Nanowells are not coated directly instead, a special poly amide disc (Solid phase) is inserted and fixed on to each NanoWell. Polyamide covalently binds the capture antigen through impregnation technology. Poly amide solid phase, having very high binding-coefficient, requires limited solid phase surface area to bind capture antigens. This makes the multiplex technology feasible wherein multiple well defined nanospots (Zone of Ag/Ab impregnation) could be accommodated onto a single poly amide disc. The stationary phase of NanoWell can accommodate up to 25 spots per polyamide disc. Capture antigens are impregnated on to a pre-defined zones/spots and that makes Multiplex testing possible. Nanoplex assays are read and interpreted by NanoScan for easy and accurate quantification and interpretation.

2. MATERIALS AND METHODS

A total of 40 blood samples were collected from different hospitals in and around Bangalore and Bagalkot city of Karnataka State. A total of 30 samples were collected from the antenatal woman with Bad Obstetric History (BOH) as the test group and 10 samples were collected from antenatal woman without any pregnancy wastage. Among the test group samples 17 were from antenatal women with history of repeated abortions, 3 from intra uterine death, 2 from intra uterine growth retardation, 4 from congenital anomalies and 4 from preterm deliveries. Cases from other causes of abortions such as Cervical incompetence, Rh incompatibility, Diabetes, Syphilis, HIV, Hepatitis were not included in this study. 2 -3 mL of venous blood is collected aseptically in serum separator vacutainers and kept in room temperature for 30 minutes to clot. Serum was separated by centrifugation and transferred to plastic aliquots. The aliquots were labeled with information of patient and were stored at -20°C until tested. Serum samples from both groups of subjects were tested using the commercial kits qualitative immune enzymatic determination of IgM and IgG antibodies for ToRCH infection.

Reagent preparation

Wash buffer

Add 2ml of 7.5 X wash buffer to 13 ml Distilled water.

Reagent 1X Blocker preparation:

Add 0.5 mL 10X blocker to 4.5 ml of 1X Wash buffer.

Sample dilution:

Serum will be diluted with 1X Blocker to give the required dilution of serum.

Preparation of 1X Secondary Antibody:

Add 40 µL 25X Secondary Antibody to 1 ml of 1X Blocker.

Preparation of 1X Detection reagent:

Add 20 µL of 50X detection reagent to 1ml of 1X Blocker.

Preparation of 1X substrate solution:

Add 50 µL of 20X substrate to 1ml 1X substrate buffer.

Test procedure

- Added 40 µL of diluted serum to the wells according to the sample layout.
- Incubated the Nanowells at 37°C for 30 minutes.

- Aspirated the solution and washed the wells 3 times with 1X wash buffer.
- Added 50 μ L freshly prepared 1x Secondary antibody.
- Incubated the Nanowells at 37°C for 30 minutes.
- Added 50 μ L of freshly prepared 1X detection reagent to each NanoWell.
- Incubated the Nanowells at 37°C for 30 minutes.
- Aspirated the solution and washed the wells 3 times with 1X wash buffer.
- Added 50 μ L of 1X substrate solution to each well.
- Incubated the Nanowells at Room temperature for 5 minutes.
- Aspirated the solution and washed the wells once with 1X wash buffer
- Dried the wells at 37°C for 30 minutes.
- Read the slide on NanoScan instrument and analyzed the results using data analysis software.

3. RESULTS

For the study of ToRCH infection, we collected 40 samples of which 30 patients were antenatal women with Bad Obstetric History (BOH). 10 samples were collected from antenatal women without any pregnancy wastage. All the samples were screened for HIV, HBsAg and HCV through Elisa method at Siddhi Diagnostics, Bagalkot using commercial kits of Meril Diagnostics (P) Ltd. All the samples were non reactive for HIV, HBsAg and HCV.

Normal ranges

NEGATIVE RANGE: 0.0 TO 0.8 IU/ML

BORDERLINE RANGE: 0.9 TO 1.3 IU/mL

POSITIVE RANGE: 1.4 TO 25 IU/mL

STRRONG POSITIVE: More than 25.1 IU/MI

Table 1: Nano Elisa titre values

		Toxoplasma		Rubella		Cytomegalo virus		HSV	
	Sample No	Ig M	IgG	Ig M	Ig G	Ig M	Ig G	Ig M	Ig G
CONROLS	1	0.1	0	0	1.5	0	0	0	0
	2	0	2.1	0	0	0.1	0	0	0
	3	0.1	0	0	0	0	0	0	0
	4	0	0	0	0.2	0	0	0	0
	5	0.1	2.4	0	0	0	0	0	0
	6	0.1	0	0	0	0	0	0	0
	7	0	0	0.1	0.1	0	0	0	0
	8	1.5	3.5	0	0	0	0	0	0
	9	0.9	4	0	0	0	0	0	0
	10	0	0.5	0	0	0	0	0	0
20-30YRS	11	0.1	0	0	0.1	0.2	7.7	0.2	7.1
	12	0.2	0	0.2	3.7	0	11.4	0	0.9
	13	0.2	10	0.2	11.1	0	0	0.3	0
	14	1	3.2	0	25	0	2.8	0	3.8
	15	0.3	3.5	0	6.2	2	8.2	0.5	2.9
	16	0.2	5.6	0.1	9.2	2	5	0	0.2
	17	2.1	9.3	2	7.7	1.5	8	0.3	6.2
	18	1	4.6	0	0	0.2	3	0	0.9
	19	0	12	0	20	0	0	0	6
	20	2.1	7	0	1.2	0.4	3	0.6	2.8
30-40 YRS	21	0.1	0	0	0.1	0.2	2	0.2	7.1

	22	0	0	0.2	0	0	0	0	0.9
	23	0.2	2	0.2	3	0	0	0.3	0
	24	0	0	0	4	0	0	0	2.2
	25	0.3	3.5	0	6.8	2	8.2	0	0
	26	0.2	0	0.1	0	2	5	0	0
	27	0	7.6	2	4.7	1.5	8	0.3	3.5
	28	1	0	0	0	0.2	3	0	0.9
	29	0	14	0	4	0	0	0	0
	30	2.1	7	0	1.2	0.4	3	0.6	1
	31	0.2	7.2	0	0	0	0	0	0
>40 YRS	32	0	0	2	4.6	0	0	0	6.6
	33	0	0	0	0	0	0	0	0
	34	0.8	2.4	0	0	0	0	0	0
	35	1.7	3.7	0	0	1.2	3.8	0	0
	36	0	0	0	0	0	0	0	0
	37	0.4	5.9	0	0	0	0	0	0
	38	0	0	0	0	0	0	0.2	2.5
	39	0.5	2.6	0	0	0	0	0	0
	40	0	3.6	0	0	0	0	0	0

Table 2: Sero-prevalence of *Toxoplasma gondii*

AGE group	Toxoplasma IgM average	Toxoplasma IgG average
Control	0.23	1.25
20-30	0.72	5.52
30-40	0.39	3.41
>40	0.36	2.54

Table 3: Seroprevalence of Rubella Virus infection

AGE group	Rubella Ig M average	Rubella Ig G average
Control	0.01	0.18
20-30	0.47	8.42
30-40	0.25	2.38
>40	0.2	0.46

Table 4: Seroprevalence of Cytomegalovirus infection

AGE group	CMV Ig M average	CMV Ig G Average
Control	0.1	0
20-30	0.63	4.91
30-40	0.35	2.92
>40	0.12	0.38

Table 5: Seroprevalence of Herpes Simplex Virus infection:

AGE group	HSV Ig M Average	HSV Ig G Average
Control	0.1	0.01
20-30	0.19	3.08
30-40	0.14	1.56
>40	0.02	0.91

4. DISCUSSION

The present study shows seropositivity rate of 8.0% for CMV specific IgM in women with BOH. In other studies seropositivity ranges from 3 to 12.9%. Seropositivity rate for HSV IgM among BOH patients in our study was 3%, whereas has been reported previously ranging from 2 to 10%

5. SUMMARY AND CONCLUSION

TORCH infections are associated with recurrent abortion, intrauterine growth retardation, intrauterine death, preterm labor, early neonatal death, and congenital malformation. Delivery wastages of earlier records and positive serological reactions during the present delivery assist management of these cases in order to reduce adverse fetal outcome. Usage of Nano-Elisa technique provides high sensitive, accurate results at economical price.

Peer-review

External peer-review was done through double-blind method.

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Conflict of Interest

The authors declare that there are no conflicts of interests.

Data and materials availability

All data associated with this study are present in the paper.

REFERENCES & NOTES

- Abida Malik, Meher Rizvi et al., Toxoplasma gondii in women with bad obstetric history and infertility: a five-year study. Asian Pac J Trop Dis 2014; 4(Suppl 1): S236-S239
- Ahmed MU. IgM and IgG antibodies specific to rubella in child bearing women. J Pak Med Assoc. 1992;42:121-2.
- Aly.E and Abo- amar (2007) characterization of a bacteriocin-like inhibitory substance produced by Lactobacillus species L.plantarum isolated from Egyptian home- made product. Science asia., 33:313-31
- Apurba S.S. , Sandhya B.K. Senthamarai S. et al., Serological Evaluation of Herpes Simplex Virus Type- 1/ Type- 2 Infections in Pregnant Women with Bad Obstetric History in a Tertiary Care Hospital, Kanchipuram. International Journal of Advanced Research (2013), Volume 1, Issue 3, 123-128
- Boonruang S, Buppasiri P. Rubella antibodies in normal pregnant women at Srinagarind hospital, KhonKaen Thailand. J Med Assoc Thai. 2005;88:455-9.
- Cao Y, Qiu L, Zhang Q. Study on the relationship between the history of abnormal pregnancy and TORCH infection in pregnant women. Zhonghua Fu Chan KeZaZhi. 1999;34:517-20.
- Chopra S, Arora U, Aggarwal A. Prevalence of IgM antibodies to Toxoplasma, rubella and cytomegalovirus infections during pregnancy. J K Sci. 2004;6:190-2.
- de Melo E J T, de Souza W. A cytochemistry study of the inner membrane complex of the pellicle of tachyzoites of Toxoplasma gondii. Parasitol Res. 1997;83:252-256.
- de Souza W, Chagas M C. Mise en évidence et structure du système microtubulaire de Toxoplasma gondii. C R Acad Sci Paris Ser D. 1972;275:2899-2901.

10. de Souza W, Souto-Pradrón T. Ultrastructural localization of basic proteins on the conoid, rhoptries and micronemes of *Toxoplasma gondii*. *Z Parasitenkd*. 1978;56:123–129.
11. Dubey JP, Beattie CP: *Toxoplasmosis of Animals and Man*. CRC Press, Boca Raton, FL, 1988 .
12. Fomda BA, Thokar MA, Farooq U, Sheikh A. Seroprevalence of rubella in pregnant women in Kashmir. *Indian J Pathol Microbiol*. 2004;47:435–7.
13. Frenkel J K. *Toxoplasma in and around us*. *BioScience*. 1973;23:343–352.
14. Guerina NG, Hsu HW, Meissner HC, Meissner HC. et al. Neonatal serologic screening and early treatment for congenital *Toxoplasma gondii* infection. *N Eng J Med*. 1994;330:1858.
15. Haider M, Rizvi M, Khan N, Malik A (2011) ISSN - 2250-1991.
16. Mehmet ÖZDEMİR, Fatma KALEM, Bahadır FEYZİOĞLU, Bülent BAYSAL. Investigation of Viral Pathogens during Pregnancy in A City Region In Turkey. *Anatol J Clin Investig* 2011;5(2):78-81
17. Mohit Bhatia, Shruthi Harle (2013) Seroprevalence of TORCH Infections and Adverse Reproductive
18. Munmun Das Sarkar, B. Anuradha, Neelam Sharma, and Rabindra Nath Roy (2012). Seropositivity of Toxoplasmosis in Antenatal Women with Bad Obstetric History in aTertiary-care Hospital of Andhra Pradesh, India. *J HEALTH POPUL NUTR* 2012 Mar;30(1):87-92ISSN 1606-0997 |
19. Namrata Kumari, Norman Morris, and Renu Dutta (2011) Is Screening of TORCH Worthwhile in Women with Bad Obstetric History: An Observation from Eastern Nepal. *J HEALTH POPUL NUTR* 2011 Feb;29(1):77-80 ISSN 1606-0997
20. Outcome in Current Pregnancy with Bad Obstetric History *J Clin Biomed Sci* 2013 ; 3 (2)
21. Padmavathy M, *Mangala Gowri, Malini J, Umapathy BL, Navaneeth BV, Paripex - Indian Journal Of Research. Volume : 2 | Issue : 11 | Nov 2013
22. Rajendra B Surpam, Usha P Kamlakar, RK Khadse, MS Qazi, Suresh V Jalgaonkar (2006) Serological study for TORCH infections in women with bad obstetric history. *J Obstet Gynecol India* Vol. 56, No. 1 : January/February 2006 Pg 41-42
23. Rema Devi, N. Sreenivas, Sayee Rajangam (2002) Bad Obstetric History and Infectious Causes. *Int J Hum Genet* 2(4): 269-271 (2002)
24. Serological study of herpes virus infection in female patients with bad obstetric history. *Biology and Medicine*, 3 (2) Special Issue: 284-290, 2011
25. Seroprevalence of Herpes Simplex Virus Type 2 (HSV 2) in Women with Bad Obstetric History. *American Journal of Dermatology and Venereology* 2013, 2(3): 31-38
26. Sucilathangam G, Anna T, Velvizhi G (2013). Seroprevalence of *Toxoplasma gondii* in Pregnant Women with Bad Obstetric History.
27. Yashodhara P, Rama Lakshmi BA, Raman L, Nadamuni Naidu. Rubella IgM positivity during pregnancy. *Indian J Med Microbiol*. 1998;16:121–2.
28. Zainab Khali, Mohamed Aljumaili et al., Seroprevalence of Herpes Simplex Virus Type 2 (HSV 2) in Women with Bad Obstetric History. *American Journal of Dermatology and Venereology* 2013, 2(3): 31-38